

SAFETY Meeting Minutes
UAMS IBC

MEETING TIME RECORDS

Meeting start time: 8/7/2026 12:00 PM

Meeting end time: 12:29 PM

Meeting type: Virtual

Name of Regular/Alternate Member	Status (Member or Alternate)	Present by Teleconference?
Ha-Neui Kim	Member	Yes
Matthew Jorgenson	Member	No
Robert Hunter	Member	No—voted via email
Kimberly Murphy	Member	Yes
Lindsey Clark	Member	Yes
James Douglas	Member	No
James Bishop	Member	No
Youssef Aachoui	Member	Yes
Jia Liu	Member	Yes
Yuet-Kin Leung	Member	Yes
Melaney Gee	Member	Yes
Mark Manzano	Member	Yes
Antino Allen	Member	No
KyoungHyun Kim	Member	Yes
James Townsend	Member	Yes
Shengyu Mu	Member	Yes
Kikumi Ono-Moore	Ex Officio	No
Zhiqiang Qin	Member	Yes
Lawson Birmingham	Member	No—voted via email
Andre Ewell	Member	Yes

QUORUM INFORMATION

Number of SAFETY members on the roster: 19

Number required for quorum: 10

Quorum: Yes

All members present via teleconference received all pertinent material before the meeting and were able to actively and equally participate in all discussions.

ATTENDANCE STATUS AND VOTING KEY	
ABSTAIN:	Present for the vote but not voting “For” or “Against.”
ABSENT:	Absent for discussion and voting for reasons other than a conflict of interest.
RECUSED:	Absent from the meeting during discussion and voting because of a conflict of interest.
SUBSTITUTION:	When regular members and their alternate(s) are listed in the ATTENDANCE table above and an alternate member serves as a substitute for the regular member this identifies the name of the alternate to indicate which individual is serving as the voting member for this vote. May be deleted if there are no substitutions.

GUEST NAMES
Lauren Grogan

Previous Meeting minutes approved: Yes

REVIEW OF SUBMISSIONS

The review and discussion of the protocols listed below included the following elements: the agents involved and their characteristics; types of manipulations planned; the source(s) and nature of the nucleic acid sequences; the host organism(s) and vector(s) to be utilized; whether expression of a foreign gene is intended and, if so, the specific protein(s) to be produced; the containment conditions to be applied, including biosafety level and any special provisions; and the relevant sections of the NIH Guidelines.

All IBC members present were reminded to identify any conflicts of interest as each registration was reviewed.

For each protocol reviewed, it was confirmed that the Principal Investigator (PI) and laboratory personnel have received appropriate training in the safe conduct of research.

De Novo Review**1. Review of SPROTO202600000046**

Title:	Tumor progression and Metastasis (BP365)
Investigator:	Md Miah
Submission ID:	SPROTO202600000046
Description:	Research Summary To uncover the molecular mechanisms by which oncogenic signaling promotes tumorigenesis and metastasis in order to identify novel therapeutic targets for cancer treatment. In Vitro Studies: Established human cancer cell lines will be genetically engineered using recombinant plasmids for gene expression or CRISPR/Cas9-mediated gene knockout to investigate genes involved in tumorigenesis and metastasis. Engineered cells will be evaluated using proliferation, migration (wound healing), invasion, western blotting, RT-qPCR, microscopy, and other molecular and biochemical assays. Recombinant human proteins expressed in Escherichia coli will be purified by affinity chromatography and analyzed by mass spectrometry to characterize protein-protein interactions and post-translational modifications. In Vivo Studies: Genetically engineered human cancer cell lines will be implanted into immunocompromised mice by subcutaneous or orthotopic injection to evaluate primary tumor growth and spontaneous metastasis, or administered by tail vein injection to assess metastatic colonization and tumor burden. Tumor growth, metastatic burden, and survival will be monitored over time. At study completion, tumors and metastatic tissues will be collected for histopathological evaluation, including hematoxylin and eosin (H&E) staining, to characterize tumor morphology and malignancy. Biosafety Considerations: The proposed research involves routine BSL-2 laboratory procedures using established human cell lines, recombinant DNA technology, CRISPR/Cas9 genome editing, recombinant protein expression in Escherichia coli, and mouse tumor models. Recombinant plasmids containing human genes of interest, selectable antibiotic resistance markers, and epitope or fluorescent tags will be used for gene expression, protein purification, and imaging.

	CRISPR/Cas9 plasmids, shRNA, and siRNA will be used to disrupt or reduce expression of target genes to investigate their roles in tumorigenesis and metastasis. Recombinant proteins will be produced in non-pathogenic <i>Escherichia coli</i> laboratory strains and purified using standard molecular biology procedures. All human cell culture procedures will be performed in a certified Class II biological safety cabinet. Chemical procedures will be conducted in a certified chemical fume hood. Biohazardous cell cultures, bacterial cultures, and recombinant materials will be decontaminated with 10% bleach or autoclaved prior to disposal in accordance with institutional biosafety procedures.
Agent Containment:	Biological Containment Levels: • Human Tumor Tissue: BSL-2 • Animal Tissue: BSL-2 • Cholera toxin: BSL-2 • Lentivirus: BSL-2 • <i>E. coli</i>: BSL-1 • MDA-MB-231 (Human TNBC cell line): BSL-2 • MCF7 Human Cell Line: BSL-2 • CFPAC-1 CLR-1918 Human Cell Line: BSL-2 • Hs578T (breast carcinoma, ER-neg) Human Cell Line: BSL-2 • HEK293E Human Cell Line: BSL-2 • HeLa cells: BSL-2 • SKBR3 Human Cell Line: BSL-2 • Human Pancreatic Duct Epithelial (HPDE) Cell Line (H6c7): BSL-2 • MCF10A Human Cell Line: BSL-2 • HCC1937: BSL-2 • BT20 Human Cell Line: BSL-2 • BxPC-3 CRL-1687 Human Cell Line: BSL-2
Applicable NIH Guidelines:	• Section III-D-4 • Section III-D-1 • Section III-D • Section III-D-3

a. **Determination:** Modifications Required

b. **Required modifications:**

Please review and respond to all comments throughout the protocol. Contact BSOs Lindsey Clark (LClark4@uams.edu) or J.C. Douglas (JDouglas@uams.edu) with any questions. Thank you!

c. **Votes:**

For: 15
Against: 0
Recused: 0
Absent: 4
Abstained: 0

Initial Protocol

2. Review of SPROTO202600000047

Title:	Glial Metabolism in Neuroinflammation
Investigator:	Cory Willis
Submission ID:	SPROTO202600000047
Description:	Glial cells are uniquely positioned within the central nervous system (CNS) to act as ‘metabolic hubs’ for the uptake, clearance, and processing of extracellular metabolic (toxic) waste. Well-regulated metabolic function is necessary for glia to sustain metabolic and biochemical processes to avoid an accumulation of neurotoxic and cytotoxic factors. CNS neuroinflammation leads to a ‘metabolic rewiring’ of glia that disrupts these metabolic and biochemical processes leading to the formation of toxic tissue microenvironments within the CNS that lead to tissue damage. However, little is known about the identity of these metabolic regulators within glia. This project will identify critical metabolic processes that are enhanced and/or disrupted during neuroinflammation leading to surrounding cell and tissue damage. Findings from this project will identify mechanistic pathways and biological targets for follow-up studies. Animal studies will include animal models of the human disease multiple sclerosis and ICV injections of inflammatory factors, including recombinant interleukin-1 beta (IL1B) and tumor necrosis factor-alpha (TNFa). Mouse CNS tissue and blood will be collected for analyses. In vitro studies will use digested neonatal mouse pup tissue to generate primary mixed glial cultures. Human glia-like cells will be generated from induced pluripotent stem cell (iPSC) lines using genetic overexpression of known transcription factors. These cells will be treated with standard inflammatory inducing agents, either lipopolysaccharide (LPS) or recombinant inflammatory factors (IL1B, TNFa, complement

	component C1q, interferon gamma) to model neuroinflammation. Additional biochemical assays and analyses will be performed to investigate how neuroinflammation affects glial cell phenotype and function. CRISPR/Cas9 interference screens using the human iPSCs will be used to identify metabolic genes critical to neuroinflammatory reactivity in glial-like cells following stimulation with inflammatory inducing agents.
Agent Containment:	Biological Containment Levels: • Lentivirus: BSL-2 • Pertussis Toxin (PTX): BSL-2 • Human Induced Pluripotent Stem Cells: BSL-2 • Adenoassociated virus AAV: BSL-2 • E. coli: BSL-2 • Heat-killed Mycobacterium tuberculosis (HKMT): BSL-2 • E. coli: BSL-2 • Animal Blood: BSL-1 • E. coli: BSL-2 • CNS tissue: BSL-1 • HEK293T Human Cell Line: BSL-2 • Animal Cells: BSL-1
Applicable NIH Guidelines:	• Section III-D-4 • Section III-D • Section III-D-3 • Section III-D-3-a

a. **Determination:** Modifications Required

b. **Required modifications:**

Please review and respond to all comments throughout the protocol. Contact BSOs Lindsey Clark (LClark4@uams.edu) or J.C. Douglas (JDouglas@uams.edu) with any questions. Thank you!

c. **Votes:**

For:	15
Against:	0
Recused:	0
Absent:	4
Abstained:	0

De Novo Review

3. Review of SPROTO202600000043

Title:	Weinkopff Biosafety Protocol 2026
Investigator:	Tiffany Weinkopff
Submission ID:	SPROTO202600000043
Description:	Cutaneous leishmaniasis has a wide spectrum of clinical manifestations, ranging from self-healing to chronic debilitating disease. The parasite is transmitted by a sand fly during a blood meal and the disease is endemic in many tropical and subtropical regions throughout the world putting 1 billion people living in endemic areas at risk for infection. Annually, about 1 million new cases are detected with major foci in Asia and South America. Presently, there is no vaccine, the drugs against the parasite are extremely toxic, and furthermore patients are often refractory to treatment. Control of the parasite is dependent upon the activation of macrophages. However, even when an appropriate adaptive immune response develops and parasites are controlled, cutaneous lesions often persist suggesting the inflammatory response can drive pathology. Thus, the goal of the proposed research will be to define the factors that control both lesion development and resolution which will be important in developing novel therapies to address the disease in patients.
Agent Containment:	Biological Containment Levels: • Animal Tissue: BSL-2 • Animal Blood: BSL-2 • Adenovirus: BSL-2 • Leishmania: BSL-2 • Leishmania: BSL-2 • Leishmania: BSL-2 • Leishmania: BSL-2 • Leishmania: BSL-2 • Leishmania: BSL-2 • Leishmania: BSL-2 • Leishmania: BSL-2 • Leishmania: BSL-2 • E. coli: BSL-2 • Leishmania: BSL-2 • HEK293T Human Cell Line: BSL-2 • Mouse Bone Marrow Macrophages: BSL-2
Applicable NIH Guidelines:	• Section III-D-1-a • Section III-E-3 • Section III-D-4 • Section III-E • Section III-D-1 • Section III-D

- a. **Determination:** Modifications Required
- b. **Required modifications:**
Please review and respond to all comments throughout the protocol. Contact BSOs Lindsey Clark (LClark4@uams.edu) or J.C. Douglas (JDouglas@uams.edu) with any questions. Thank you!
- c. **Votes:**

For:	15
Against:	0
Recused:	0
Absent:	4
Abstained:	0

REVIEW OF OTHER AGENDA ITEMS

No other items reviewed.